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The Equilibrium between [CpCr(CO)₃]₂ and CpCr(CO)₃. The Thermodynamics of Cr-Cr Single Bond Cleavage

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There is NMR,¹ ESR,² and electrochemical² evidence for the reversible thermal dissociation of [CpCr(CO)₃]₂ in solution (reaction 1). We wish to report a quantitative study of this equi-

$$[CpCr(CO)_3]_2 \rightleftharpoons 2CpCr(CO)_3 \tag{1}$$

librium which demonstrates that the equilibrium concentration of radical 2 is much higher than previously supposed. We also report the first infrared spectrum of 2 in solution.

Solutions of 1 have a strong UV absorbance that is associated with the Cr–Cr bond $[\lambda_{max} = 450 \text{ nm}, 1(\sigma^2) \rightarrow 1(\sigma\sigma^*)]^{3,4}$ Upon dilution, this absorbance decreases more than expected from Beer's law behavior. If the diluted solution is reconcentrated, the original intensity of the 450-nm band is restored. Spectra recorded at different concentrations and scaled to adjust for concentration show an isosbestic point at 370 nm. These observations are consistent with a concentration dependent equilibrium between 1 and a species with no Cr-Cr bond.

The UV spectrum of 1 is temperature dependent. Increasing the temperature from -65 °C to +95 °C causes a reversible reduction in the intensity of the 450-nm band (Figure 1). This dramatic temperature dependence is consistent with the reversible formation of 2 (reaction 1). At temperatures below -35 °C, where the equilibrium is shifted entirely to 1, the spectra have an isosbestic point at 456 nm (Figure 1 insert). The isosbestic behavior of 1 can be interpreted as a temperature dependent equilibrium between two rotational isomers with similar but not identical UV spectra (Figure 2).^{1,5} [CpMo(CO)₃]₂, which is also known to be an equilibrium mixture of two rotomers,⁶ exhibits similar isosbestic behavior (toluene solution, -35 °C to +45 °C, $\lambda_{max} = 390$ nm, isosestic point = 400 nm). The spectra shown in Figure 1 can be fit to the equilibrium model of reaction 1 to give accurate values of ΔH , ΔS , ϵ_1 , and ϵ_2 (Table I).⁷ Experiments in THF give similar results.



Figure 1. UV spectra of 1 in toluene (0.0020 M, 0.10 cm cell) from -65 °C to 95 °C (20 °C increments). Insert is -65 °C to 5 °C (10 °C increments). Spectra are corrected for the temperature dependence of the solvent density. The discontinuities at 360, 490, and 660 nm are artifacts.



Figure 2. Equilibrium between anti dimer, gauche dimer, and monomer.

Table I. Thermodynamic Parameters^a for the $1 \Rightarrow 2$ Equilibrium

method	solvent	ΔH	ΔS	ΔG_{300}
UV ^b IR	toluene	15.8 (0.4) 14.8 (0.4)	37.1 (1.4) 34.3 (1.3)	4.6
	toluene			4.6
UV۴	UV ^c THF IR THF			4.5
IR				4.7

^a ΔH and ΔG in kcal mol⁻¹; ΔS in cal mol⁻¹ K⁻¹; figures in parentheses are esd's. ${}^{b}\epsilon_{1}$ (456 nm) = 17740 (960); ϵ_{2} (456 nm) = 328 (106). ${}^{c}\epsilon_{1}$ (446 nm) = 18600 (990); ϵ_{2} (446 nm) = 180 (150).

IR spectra of 1 in solution show direct evidence of 2. The complex solution spectrum of 1 has been attributed to an equilibrium mixture of anti and gauche rotometers (Figure 2, Figure 3a).⁵ However, the concentration dependence of the spectrum (Figure 3a,b) can only be accounted for by an additional intermolecular equilibrium (i.e., reaction 1). Subtracting an appropriately scaled spectrum of a concentrated sample from the spectrum of a less concentrated sample gives a spectrum of 2, the species favored at lower concentration (Figure 3c). This spectrum can be compared with the spectrum of 2 produced by photolysis of CpCr(CO)₃H in a CO matrix ($\nu_{CO} = 1986, 1910, 1902 \text{ cm}^{-1}$).⁸ The subtraction factor SF used to obtain Figure 3c can be used to calculate the equilibrium constant of reaction 1.9 This value

⁽¹⁾ Adams, R. D.; Collins, D. E.; Cotton, F. A. J. Am. Chem. Soc. 1978, 96, 749-754.

⁽²⁾ Madach, T.; Vahrenkamp, H. Z. Naturforsch., B: Anorg. Chem., Org. Chem. 1978, 33, 1301-1303.

⁽³⁾ Madach, T.; Vahrenkamp, H. Z. Naturforsch., B: Anorg. Chem., Org. Chem. 1979, 34, 573-578. (4) Meyer, T. J.; Caspar, J. V. Chem. Rev. 1985, 85, 187-218.

⁽⁵⁾ Hackett, P.; O'Neill, P. S.; Manning, A. R. J. Chem. Soc., Dalton

⁽⁵⁾ Hackett, P.; O'Neill, P. S.; Manning, A. R. J. Chem. Soc., Datton Trans. 1974, 1625-1627. (6) Adams, R. D.; Cotton, F. A. Inorg. Chim. Acta 1973, 7, 153-156. (7) Starting with the definition of the equilibrium constant for reaction 1, $K_{eq} = [2]^2/[1]$, the following model was derived: absorbance $= \epsilon_1 [1] + \epsilon_2 [12]; [1] = \{[Cr_{tot}] - [2]\}/2; [2] = \{-K_{eq} + (K_{eq}^2 + 8K_{eq}[Cr_{tot}])^{1/2}]/4; K_{eq} = e^{-\Delta G/RT}$, $\Delta G = \Delta H - T\Delta S; [Cr_{tot}] = \{[Cr_{25}]/d_{25}]/d_0 + AT_c + BT_c^2 + CT_c^3\}$ where ϵ_1, ϵ_2 = extinction coefficients of 1 and 2 at the isosbestic frequency; 1 = UV cell pathlength (cm); $[Cr_{tot}] =$ the total concentration of Cr atoms in solution; T, T_c = temperature in K and °C; $[Cr_{25}] = Cr$ atom concentration of the starting solution at 2 S°C, d, d_{es} = solvet densities at 0 °C and 25 of the starting solution at 25 °C; d_0 , d_{25} = solvent densities at 0 °C and 25 °C; A, B, C = solvent thermal expansion coefficients. The data were least-squares fit to this model by using SAS/ETS software with Gauss-Newton minimization.

⁽⁸⁾ Mahmoud, K. A.; Rest, A. J.; Alt, H. G. J. Chem. Soc., Chem. Com-mun. 1983, 1011-1013.

mun. 1983, 1011–1013. (9) The conditions for obtaining a null of 1 in a spectral substraction are $[1]_a - SF[1]_b = 0$. [1] can be expressed as a function of K_{eq} and $[Cr_{tot}]$: [1] $= [4[Cr_{tot}] + K_{eq} - (8[Cr_{tot}]K_{eq} + K_{eq}^2)^{1/2}]/8$. Substitution of this equation into the spectra subtraction equation gives an equation of the form $f(K_{eq}, -[Cr_{tot}]_a, [Cr_{tot}]_b, SF) = 0$. The value if K_{eq} was determined by an iterative root finding program on a pocket calculator.



Figure 3. FTIR spectra of 1 in THF: (a) 0.005 M, (b) 0.0005 M, (c) subtraction B-(SF)A.

is in good agreement with the results of UV experiments, thus confirming our equilibrium model (Table I). As the temperature is increased, IR spectra show a shift in the equilibrium toward 2. At 92 °C, the equilibrium is shifted almost entirely to 2, and the IR spectrum resembles Figure 3c.

The thermodynamic parameters indicate an extremely weak Cr-Cr bond for 1. The amount of radical 2 present in solutions of 1 at room temperature is much greater than previously estimated.² For example, a 0.01 M solution of 1 at 25 °C is about 10% dissociated to 2. The large positive value of ΔS and its insensitivity to solvent (toluene versus THF) both argue against any direct participation of solvent in reaction 1. Similar values of ΔS have been reported for Fe-Fe bond cleavage in [(allyl)- $Fe(CO)_{3}_{12}^{10}$ Upon the basis of the heat of hydrogenation of 1 and an estimate of the Cr-H bond strength, Hoff had previously estimated the enthalpy of reaction 1 to be 12.7 kcal/mol.¹¹ Considering the approximation use by Hoff, his estimate is in good agreement with our results.

Upon the basis of the known lability of 17-electron radicals,¹² solutions of 1/2 are expected to react readily with donor ligands. Indeed, the carbonyl ligands of 1 exchange with ¹³CO at 25 °C and subatmospheric pressure! Reaction with Me₃CNC yields the stable monomeric 17-electron complex CpCr(CO)₂(CNCMe₃).¹³

Registry No. [CpCr(CO)₃]₂, 12194-12-6; CpCr(CO)₃, 12079-91-3; CpCr(CO)₂(CNCMe₃), 112043-97-7; Me₃CNC, 7188-38-7.

(10) Muetterties, E. L.; Sosinsky, B. A.; Zamaraev, K. I. J. Am. Chem. Soc. 1975, 97, 5299-5300

(11) Landrum, J. T.; Hoff, C. D. J. Organomet. Chem. 1985, 282, 215-224.

(12) Therien, M. J.; Ni, C.-L.; Anson, F. C.; Osteryoung, J. C.; Trogler, W. C. J. Am. Chem. Soc. **1986**, 108, 4037–4042. (13) $CpCr(CO)_2(CNCMe_3)$. A solution of Me₃CNC (0.083 g, 1.00 mmol) in 25 mL of pentane was added dropwise to a rapidly stirred suspension of $[CpCr(CO)_3]_2$ (0.201 g, 0.50 mmol) in 75 mL of pentane over a period of 30 min. After having been stirred an additional 10 min, the mixture was filtered and cooled to -40 °C to give dark red-brown needles, 0.176 g (69%): IR (pentane solution, cm⁻¹) 2096 (w), 2071 (ww), 1938 (vs), 1842 (s); ESR (3-methylpentane, -80 °C) 2.0423. Anal. Calcd for C₁₂H₁₄CrNO₂: C, 56.25; H, 5.51; Cr, 20.3; N, 5.47. Found: C, 56.19; H, 5.46; Cr, 19.6; N, 5.63.

DNA Strand Scission by (-)-Epicatechin and Procyanidin B₂

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In recent years, considerable effort has been made to identify and characterize molecules capable of mediating nucleic acid strand scission. These include natural¹ and synthetically derived² products as well as known DNA binders that have been modified with adjuvants capable of "sequence-neutral" cleavage.³ Interestingly, in spite of the natural origin of many of these species, we are unaware of any systematic effort to identify additional natural products that mediate nucleic acid strand scission. Reported herein is the identification of two plant-derived natural products that mediate DNA strand scission at micromolar concentrations.

(-)-Epicatechin $(1a)^4$ and (-)-epicatechin- $[4\beta-8]$ -(-)-epicatechin (1b) (procyanidin B_2^{5}) were isolated from an extract of *Celastrus*



(1) See e.g.: (a) Sausville, E. A.; Peisach, J.; Horwitz, S. B. Biochemistry 1978, 17, 2740. (b) Chang, C.-H.; Meares, C. F. Biochemistry 1982, 21, 6332. (c) Suzuki, H.; Kirino, Y. Tanaka, N. J. Antibiot. (Tokyo) 1983, 36, (d) Aft, R. L.; Mueller, G. C. J. Biol. Chem. 1983, 258, 12069. (e)
 Hurley, L. H.; Reynolds, V. L.; Swenson, D. H.; Petzold, C. L.; Scahill, T. A. Science (Washington, D.C.) 1984, 226, 843. (f) Ueda, K.; Morita, J.; Komano, T. Biochemistry 1984, 23, 1634. (g) Povirk, L. F.; Goldberg, I. H. Biochemistry 1984, 23, 6304. (h) Eliot, H.; Gianni, L.; Myers, C. Biochem istry 1984, 23, 928. (i) Hecht, S. M. Acc. Chem. Res. 1986, 19, 383. (j) Kuwahara, J.; Suzuki, T.; Funakoshi, K.; Sugiura, Y. Biochemistry 1986, 25, 1216

1216.
(2) (a) Que, B. G.; Downey, K. M.; So, A. G. Biochemistry 1980, 19, 5987.
(b) Marshall, L. E.; Graham, D. R.; Reich, K. A.; Sigman, D. S. Biochemistry 1981, 20, 244. (c) Hertzberg, R. P.; Dervan, P. B. J. Am. Chem. Soc. 1982, 104, 313. (d) Hënichart, J.-P.; Houssin, R.; Bernier, J.-L.; Catteau, J.-P. J. Chem. Soc., Chem. Commun. 1982, 1295. (e) Sugiura, Y.; Suzuki, T.; Otsuka, M.; Kobayashi, S.; Ohno, M.; Takita, T.; Umezawa, H. J. Biol. Chem. 1983, 258, 1328. (f) Wong, A.; Huang, C.-H.; Crooke, S. T. Biochemistry 1984, 23, 2939. (g) Wong, A.; Huang, C.-H.; Crooke, S. T. Biochemistry 1984, 23, 2946. (h) Barton, J. K.; Raphael, A. L. J. Am. Chem. Soc. 1984, 106, 2466. (i) Barton, J. K.; Raphael, A. L. D. Cham. Soc. 1984. Soc. 1984, 106, 2466. (i) Barton, J. K.; Raphael, A. L. Proc. Natl. Acad. Sci.
 U.S.A. 1985, 82, 6460. (j) Müller, B. C.; Raphael, A. L.; Barton, J. K. Proc.
 Natl. Acad. Sci. U.S.A. 1987, 84, 1764. (k) Sigman, D. S. Acc. Chem. Res. 1986, 19, 180.

(3) (a) Lown, J. W.; Joshua, A. V. J. Chem. Soc., Chem. Commun. 1982, 1298.
(b) Schultz, P. S.; Dervan, P. B. Proc. Natl. Acad. Sci. U.S.A. 1983, 80, 6834.
(c) Schultz, P. S.; Dervan, P. B. J. Am. Chem. Soc. 1984, 105, 7748.
(d) Bowler, B. E.; Hollis, L. S.; Lippard, S. J. J. Am. Chem. Soc. 1984, 106, 6102.
(e) Dreyer, G. B.; Dervan, P. B. Proc. Natl. Acad. Sci. U.S.A. 1985, 82, 963.
(f) Chu, C. F.; Orgel, L. E. Proc. Natl. Acad. Sci. U.S.A. 1985, 82, 963.
(g) Vlassov, V. V.; Gaidamakov, S. A.; Gorn, V. V.; Grachev, S. A. FEBS Lett. 1985, 182, 415.
(h) Youngquist, R. S.; Dervan, P. B. J. Am. Chem. Soc. 1985, 107, 5528.
(i) Baker, B. F.; Dervan, P. B. J. Am. Chem. Soc. 1985, 32, 291.
(k) Zarytova, V. F.; Kutyavin, I. V.; Sil'nikov, V. N.; Shishkin, G. V. Bioorg. Khim. 1986, 12, 911.
(l) Iverson, B. L.; Dervan, P. B. J. Am. Chem. Soc. 1987, 109, 1241.
(m) Biodot-Forget, M.; Thuong, N. T.; Chassignol, M.; Hélene, C. C. R. Acad. Sci. Ser. 3, 1986, 302, 75.
(4) (a) Weinges, K.; Bahr, W.; Ebert, W.; Goritz, K.; Marx, H.-D. In Fortschritte der Chimie Organischer Naturstoffe; Zechmeister, L., Ed.; (3) (a) Lown, J. W.; Joshua, A. V. J. Chem. Soc., Chem. Commun. 1982, Fortschritte der Chimie Organischer Naturstoffe; Zechmeister, L., Ed.; Springer-Verlag: New York, 1969; p 159. (b) Freundenberg, K.; Weinges, K. In The Chemistry of Flavanoid Compounds; Gessman, T. A., Ed.; Pergamon: Oxford, 1962; p 197.

0002-7863/88/1510-0644\$01.50/0 © 1988 American Chemical Society